

Preparation and Enzyme-based Structure Determination of
 $2^A, 6^E$ -Bis(O-disulfonyl)- β -cyclodextrin

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2,6-di-O-Disulfonates were prepared by the reaction of 2-O- (mesitylsulfonyl)- β -cyclodextrin with mesitylsulfonyl chloride in pyridine. Each regioisomer was isolated and the $2^A, 6^E$ -isomer was structurally determined through enzymatic hydrolysis by Taka amylase A and bacterial saccharifying amylase.

Sulfonylation of two hydroxyl groups of cyclodextrins has been extensively studied since the disulfonylated cyclodextrins are starting compounds for the preparation of enzyme mimics.¹⁾ However, most of the studies are limited to sulfonylation of two primary hydroxyl groups (6-OHs)²⁾ or two secondary hydroxyl groups (2-OHs³⁾ or 3-OHs⁴⁾), i.e. disulfonylation on the same-side of cyclodextrins.

Recently, we reported the preparation of 3^A -O-(β -naphthylsulfonyl)- 6^X -O-mesitylsulfonyl- α -cyclodextrins (X = A, B, C, D, E, and F) and their structure determination through chemical con-

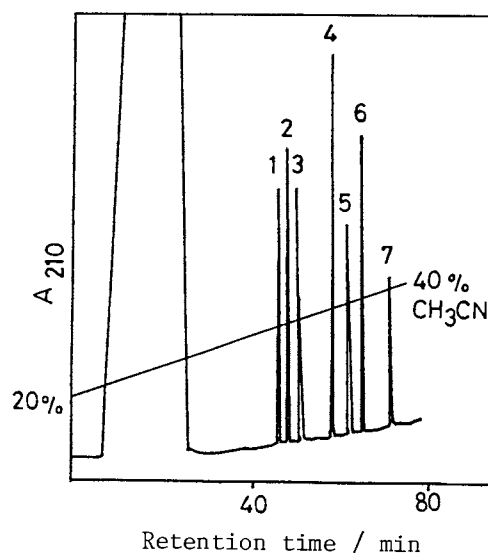


Fig. 1. Reverse-phase HPLC of the mixture of disulfonates 1-7. An gradient elution of aqueous CH₃CN was applied.

versions to $3^A, 6^A; 3^X, 6^X$ -dianhydro- α -cyclodextrins ($X = B, C, \text{ and } D$).⁵⁾

However, this method for structure determination cannot be applied to the case of $2^A, 6^X$ -disulfonates of cyclodextrins. We describe here for the first time the preparation, isolation, and structure determination of a β -cyclodextrin (2) which has sulfonates of 2^A -OH and 6^E -OH.

A reaction of 2-O-mesitylsulfonyl- β -cyclodextrin (550 mg) with mesitylsulfonyl chloride (1.37 g) in pyridine (20 mL) at room temperature for 15 min followed by usual work-up procedures gave a mixture of seven disulfonates 1-7 [the numbers of the compounds are given in the order of in-

creasing retention time in reverse-phase HPLC (Fig. 1)]. Separation by use of the reverse-phase column gave a mixture (65 mg, 10.4%) of 1, 2, and 3 and each of 4-7 (2-3%). Each component (1; 13.0 mg, 2.1%, 2; 9.8 mg, 1.6%, 3; 8.2 mg, 1.3%) was isolated from the mixture by preparative HPLC with elution of 30% aqueous CH_3CN at flow rate 1.0 mL/min. The fast atom bombardment mass spectra (FABMS) and ^1H and ^{13}C NMR spectra demonstrated that they were 2,6-di-O-disulfonates of β -cyclodextrin. Therefore, each of the seven products 1-7 must be one of the possible seven isomers, $2^A, 6^X$ -bis(O-mesitylsulfonyl)- β -cyclodextrins ($X = A, B, C, D, E, F, \text{ and } G$). Since the regiochemistry of the compounds cannot be determined by the spectral data mentioned above, we adopted an enzyme-based method for structure determination.

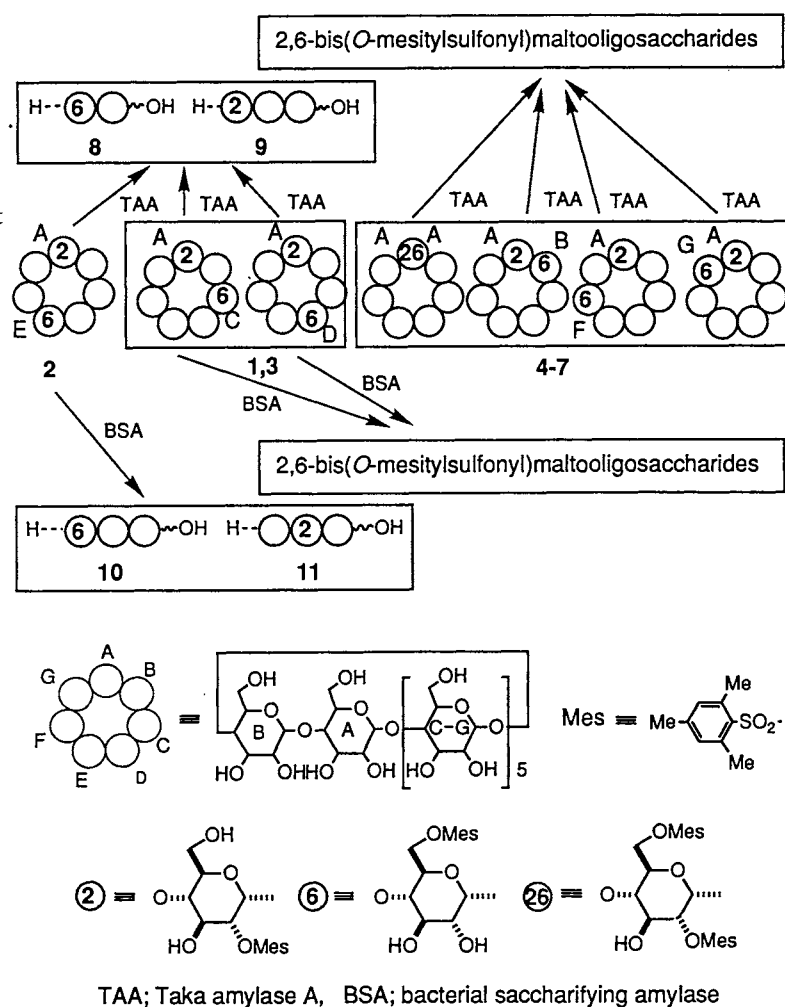


Fig. 2. Enzymatic hydrolysis of $2^A, 6^X$ -bis(O-mesitylsulfonyl)- β -cyclodextrin.

Each of enzymatic hydrolysis of **1**, **2**, and **3** by Taka amylase A (hereafter abbreviated as TAA) gave a mixture of 6'-O-(mesitylsulfonyl)maltose **8** and 2''-O-(mesitylsulfonyl)-maltotriose **9** (Fig. 2). Similar enzymatic hydrolysis of **4-7** by TAA gave bis(mesitylsulfonyl)maltotetraose different from each other. The former results show that one of **1-3** is one of the 2^A,6^D-, 2^A,6^E-, and 2^A,6^C-disulfonates, but cannot discriminate among them. The latter results confirm only that one of **4-7** is one of the 2^A,6^F-, 2^A,6^B-, 2^A,6^A-, and 2^A,6^G-isomers. Both results are in accord with the expectation based on the action pattern of TAA in the hydrolysis of 2-O-arylsulfonyl- β -cyclodextrin and 6-O-arylsulfonyl- β -cyclodextrin,⁶⁾ except for the case of the 2^A,6^A- and 2^A,6^G-isomers which are expected to give bis(mesitylsulfonyl)maltotriose.

In this situation, we disclose that an alternative enzymatic hydrolysis can differentiate the 2^A,6^E-isomer from the 2^A,6^D- and 2^A,6^C-isomers effectively. Preliminarily, we already found that bacterial saccharifying amylase (hereafter abbreviated as BSA) hydrolyzed 6-O-mesitylsulfonyl- β -cyclodextrin and 2-O-mesitylsulfonyl- β -cyclodextrin to give 6''-O-(mesitylsulfonyl)maltotriose (**10**) and 2'-O-(mesitylsulfonyl)maltotriose (**11**), respectively.⁷⁾ These findings suggest that only the 2^A,6^E-isomer gives a mixture of **10** and **11** and that each of the other two isomers affords an oligosaccharide having two mesitylsulfonyl groups.

BSA (0.1 mg) was added to an ice-cooled solution of **2** (5.0 mg) in water (1 mL). The reaction mixture was kept at room temperature for 4 d and then in boiling water for 10 min. After filtered, the solution was analyzed by reverse-phase HPLC with gradient elution from 20% aqueous CH₃CN (30 mL) to 50% aqueous CH₃CN (30 mL): the retention time (flow rate; 0.5 mL/min), **10**; 17 min, **11**; 39 min. The sulfonates were isolated by reverse-phase HPLC and analyzed by FABMS: *m/z* **10** 687 (M+H⁺), **11** 687 (M+H⁺), 709 (M+Na⁺), 725 (M+K⁺). By comparing their HPLC retention times of **10** and **11** with those of the corresponding authentic compounds, **10** and **11** were determined to be 6''-O-(mesitylsulfonyl)maltotriose and 2'-O-(mesitylsulfonyl)maltotriose, respectively. However, **1** and **3** gave bis(mesitylsulfonyl)oligosaccharides as expected respectively.

The finding that **2** was enzymatically hydrolyzed to **10** and **11** indicates that **2** was either the 2^A,6^E- or the 2^A,6^F-disulfonate. Since these two disulfonates can be differentiated by the TAA hydrolysis as mentioned above, **2** is assigned to 2^A,6^E-bis(O-mesitylsulfonyl)- β -cyclodextrin.

In conclusion, combination of TAA and BSA hydrolysis is a very useful and convenient method for the structure determination of cyclodextrin derivatives and the 2^A,6^E-disulfonate is the first cyclodextrin that possesses both 2-O- and 6-O-sul-

fonates and is structurally determined unequivocally.

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